Multiple Myeloma

The expression of PRDI-BF1 β isoform in multiple myeloma plasma cells

The PRDM1 gene, a master regulator of plasma cells (PC), can generate two transcription factor isoforms: PRDI-BF1 α and PRDI-BF1 β . The present study shows that purified human normal PC have a significantly lower level of PRDI-BF1 β expression than that in tumoral PC isolated from multiple myeloma (MM) (0.06±0.01 and 0.25±0.05, respectively; p<0.001). The role of this finding in MM is discussed.

Haematologica 2006; 91:1579-1580

Human positive regulatory domain I binding factor 1 (PRDI-BF1 or BLIMP-1) is a transcription factor that has been demonstrated to act as a master regulator required and sufficient for the generation and for the prolonged maintenance of plasma cells (PC).1 PRDI-BF1 essentially functions as a repressor, causing exit from cell cycling and the extinction of the expression of several genes critical for B-cell development at earlier stages.² The PRDM1 gene, which codifies for PRDI-BF1, contains an alternative promoter capable of generating a PRDI-BF1 deleted protein (called PRDI-BF1ß), which lacks 101 amino acids comprising most of the regulatory domain (Figure 1A). PRDI-BF1ß has been detected in relevant quantities in multiple myeloma (MM) cell lines (U266 and NCI-H929).³ Since this molecule contains the DNA-binding domain but bears a disrupted regulatory domain, PRDI-BF1 β might behave as an inhibitor of functional PRDI-BF1, called PRDI-BF1 α . We decided to compare, using real time polymerase chain reaction (RT-PCR), the occurrence of $PRDI-BF1\alpha$ and $PRDI-BF1\beta$ in human MM cell lines (U266 and NCI-H929, from ECACC, Salisbury, UK), MM patients' tumoral PC (n=17) freshly purified by immunomagnetic separation with monoclonal anti-CD138-coated microbeads (Miltenyi-Biotec, Auburn, CA, USA) and normal human PC (n=11) purified from either bone marrow, tonsil or colon lamina propria, as previously described.⁴ In order to perform a relative quantitative RT-PCR analysis of both factors, two different oligonucleotides pairs were used, one detecting both the α - and β -isoforms, and the other recognizing only the PRDI-BF1β isoform (Figure 1B). Total RNA was extracted and reverse transcribed from purified PC using RNA isolation and transcriptor kits from Roche (Barcelona, Spain). Primer mixes and TaqMan probes, assay references Hs00153357_m1 for PRDI-BF1(α and β) and

Figure 2 (right). Expression of PRDI-BF1 transcripts measured by real-time PCR. All transcript expression results are calculated relative to β -actin and are expressed in arbitrary units. The Mann-Whitney U test was used to determine the statistical differences. A. PRDI-BF1 α transcript expression in MM cell lines (U266 and NCI-H929), in human normal PC (N-PC) and in MM patients` PC (MM-PC). The data were obtained by subtracting the value observed with Probe-2 (specific for the β isoform) from values observed with Probe-1 that detected α and β isoforms in each experiment. B. PRDI-BF1 β transcript expression levels obtained with Probe-2. C) Ratio of PRDI-BF1 α /PRDI-BF1 β considering the results of expression levels obtained from A) and B). The results represent the mean±s.e.m. of 4, 3, 11 and 17 different experiments for U266 and NCI-H929 cell lines, N-PC and MM-PC, respectively. *=p<0.05 and **=p<0.01 compared to U266. NS: not significant.

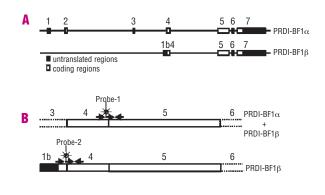
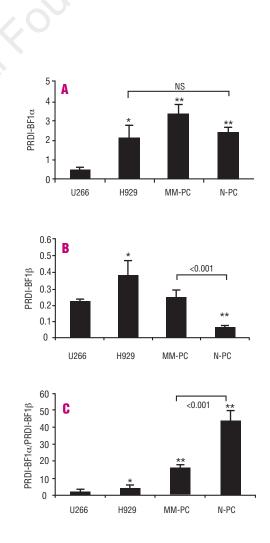


Figure 1. PRDM1 gene organization and primer localizations. A. Schematic exon-intron distribution of the PRDM1 gene showing the composition of the PRDI-BF1 α and PRDI-BF1 β isoform exons. B. Locations of probes and of primer pairs on cDNA used for detecting the two transcripts. Probe-1 hybridizes on the sequence junction between exon-4 and exon-5 (common to both isoforms) and does not discriminate between the two transcript isoforms. Probe-2 (5'-(FAM)-CTCTGGAATAGATCTTTTC-(TAMRA)-3') is a specific probe for detecting the PRDI-BF1 β isoform which hybridizes on the sequence junction between exon-1 and exon-2 for the PRDI-BF1 β isoform. The forward primer sequence was 5'-CCGAACAAGACGATAAAACTGA-3' and the reverse primer sequence 5'-CCGTCAATGAAGTGGTGAAGCT-3'.



Hs99999903_m1 for β -actin (used as an internal reference), were obtained from Applied Biosystems (Foster City, CA, USA). For PRDI-BF1ß specific detection, a probe was synthesized using the Assays-by-Design Service (Applied Biosystems). The quantity of PRDI-BF1 α was obtained by subtracting the PRDI-BF1 β value from the quantity obtained for both isoforms together. The relative quantitative values were calculated using the $2^{-\Delta\Delta CT}$ method.5

Figure 2A shows that PRDI-BF1 α was expressed at considerable levels in all PC samples tested and no significant differences were found in the PRDI-BF1 α expression between the MM cell line NCI-H929, normal PC and MM patients PC $(2.10\pm0.66; 2.38\pm0.26; 3.32\pm0.52,$ respectively: arbitrary units, mean±s.e.m.). The reduced level of PRDI-BF1 α observed in the U266 cell line (0.5±0.11; mean±s.e.m.) has been previously reported in a study using a ribonuclease protection assay.³ The relatively high expression of the α -isoform in all cell types tested could be expected bearing in mind that its protein product, PRDI-BF1 α , is strictly required for commitment to the PC fate.^{1,6-8} When the expression level of the PRDI-BF1 β isoform was tested and compared in the same PC populations (Figure 2B), significantly greater differences were observed. Interestingly, normal human PC had a markedly lower transcript level for this factor (0.06 ± 0.01) than did the other three MM PC populations $(0.23\pm0.01,$ 0.38 ± 0.09 and 0.25 ± 0.05 , for U266 and NCI-H929 MM cell lines, and MM patients' PC, respectively; mean \pm s.e.m.). These data suggest that the level of PRDI-BF1 β expression could be a feature distinguishing between malignant and normal PC. For this reason we decided to examine the ratio of PRDI-BF1 α /PRDI-BF1 β transcript levels in the different PC under study. As shown in Figure 2C, MM cell lines had very low ratios (2.26±0.56 and 5.28±0.35 for U266 and NCI-H929, respectively), a result that was due to the high level of PRDI-BF1 β expression. In contrast, normal PC had a tenfold higher PRDI-BF1α/PRDI-BF1β ratio (43.68±5.89). The ratios obtained in normal PC were more than three times higher than those of MM patients' PC (16.37 ± 1.04). The physiological significance of the differential expression of the two PRDI-BF1 isoforms by PC remains to be elucidated. In this regard, other PRDM family members have also been shown to be expressed in two different isoforms, the deleted one acting as a negative regulator of the other.9 In addition, the tumorigenic capacity of a defective PRDI-BF1 α has been recently observed in human diffuse large B-cell lymphoma.10 Therefore, it is conceivable that the overexpression of PRDI-BF1^β detected in MM patients' PC could contribute to progression into the tumoral state.³

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Funding: This work was supported by Grants G03/136 and PI052357 from Fondo de Investigaciones Sanitarias from the Ministerio 'de Sanidad y Consumo, and Grant 201/03 from Junta de Andalucía of Śpain.

Key words: multiple myeloma, PRDI-BF1/Blimp-1, real-time quantitative polymerase chain reaction.

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References

- Shapiro-Shelef M, Lin KI, Savitsky D, Liao J, Calame K. Blimp-1 is required for maintenance of long-lived plasma cells in the bone marrow. J Exp Med 2005;202:1471-6.
 Lin KI, Angelin-Duclos C, Kuo TC, Calame K. Blimp-1-dependent repression of Pax-5 is required for differentiation of B cells to immunoglobulin M-secreting plasma cells. Mol Cell Biol 2002;22:4771-80.
 Course L Bairo C, Check N, Sata F, Wright KL, Identification of
- 3. Gyory I, Fejer G, Ghosh N, Seto E, Wright KL. Identification of a functionally impaired positive regulatory domain I binding factor 1 transcription repressor in myeloma cell lines. Immunol. 2003;170:3125-33.
- Medina F, Segundo C, Campos-Caro A, Gonzalez-Garcia I, Brieva JA. The heterogeneity shown by human plasma cells from tonsil, blood, and bone marrow reveals graded stages of increasing maturity, but local profiles of adhesion molecule expression. Blood 2002;99:2154-61. 5. Livak KJ, Schmittgen TD. Analysis of relative gene expression
- data using real-time quantitative PCR and the 2(-ΔΔC^T) Method. Methods 2001;25:402-8.
 Shapiro-Shelef M, Lin KI, McHeyzer-Williams LJ, Liao J, McHeyzer-Williams MG, Calame K. Blimp-1 is required for
- the formation of immunoglobulin secreting plasma cells and pre-plasma memory B cells. Immunity 2003;19:607-20. 7. Turner CA Jr, Mack DH, Davis MM. Blimp-1, a novel zinc fin-
- ger-containing protein that can drive the maturation of B lymphocytes into immunoglobulin-secreting cells. Cell 1994; 77:297-306.
- Angelin-Duclos C, Cattoretti G, Lin KI, Calame K. Commitment of B lymphocytes to a plasma cell fate is associ-ated with Blimp-1 expression in vivo. J Immunol 2000; 165:5462-71
- Chadwick RB, Jiang GL, Bennington GA, Yuan B, Johnson CK, Stevens MW, et al. Candidate tumor suppressor RIZ is fre-quently involved in colorectal carcinogenesis. Proc Natl Acad Sci USA 2000;97:2662-7
- Tam W, Gomez M, Chadburn A, Lee JW, Chan WC, Knowles DM. Mutational analysis of PRDM1 indicates a tumor suppressor role in diffuse large B-cell lymphomas. Blood 2006; 107:4090-100.